This article was downloaded by:

On: 26 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



# Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: <a href="http://www.informaworld.com/smpp/title~content=t713597286">http://www.informaworld.com/smpp/title~content=t713597286</a>

# Nucleoside Analogues on the Basis of 4(R),5(R)-Dihydroxymethyl-2-methyl-1,3-dioxolane

Sergey N. Mikhailov<sup>a</sup>; Ekaterina V. Efimtseva<sup>a</sup>; Sergey V. Meshkov<sup>b</sup>; Earl R. Kern<sup>c</sup>
<sup>a</sup> Engelgardt Institute of Molecular Biology, the Russian Academy of Sciences, Moscow, Russia <sup>b</sup>
Semyonov Institute of Chemical Physics, the Russian Academy of Sciences, Moscow, Russia <sup>c</sup>
Department of Pediatrics, University of Alabama, Birmingham, Alabama, USA

To cite this Article Mikhailov, Sergey N. , Efimtseva, Ekaterina V. , Meshkov, Sergey V. and Kern, Earl R.(1994) 'Nucleoside Analogues on the Basis of 4(R),5(R)-Dihydroxymethyl-2-methyl-1,3-dioxolane', Nucleosides, Nucleotides and Nucleic Acids, 13: 1, 615 — 623

To link to this Article: DOI: 10.1080/15257779408013267 URL: http://dx.doi.org/10.1080/15257779408013267

## PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

# NUCLEOSIDE ANALOGUES ON THE BASIS OF 4(R),5(R)-DIHYDROXYMETHYL-2-METHYL-1,3-DIOXOLANE

Sergey N. Mikhailov<sup>1</sup>, Ekaterina V. Efimtseva<sup>1</sup>, Sergey V. Meshkov<sup>2</sup> and Earl R. Kern<sup>3</sup>

<sup>1</sup> Engelgardt Institute of Molecular Biology, the Russian Academy of Sciences, Vavilov str. 32, Moscow 117984, Russia;

ABSTRACT. New nucleoside analogues on the basis of 4(R), 5(R)-dihydroxymethyl-2-methyl-1, 3-dioxolane 2 have been prepared. Alkylation of thymine, adenine and N<sup>2</sup>-palmitoyl-guanine with 2-bromomethyl-4(R), 5(R)-dibenzyloxymethyl-1, 3-dioxolane followed by separation of regio isomers by adsorption chromatography and deprotection yielded the desired chiral nucleoside analogues. The structures of thus prepared compounds were confirmed by UV and PMR spectroscopy. The obtained compounds 11 have no anti-HIV and antiherpetic activity and are not cytotoxic.

#### INTRODUCTION

The discovery of 2',3'-dideoxynucleosides  $^{1,2}$  as potent inhibitors of the HIV virus has stimulated intensive efforts in this field (for a recent review see  $^3$ ). Recent studies of a number of nucleosides in which the ribose moiety was replaced by five membered rings such as dioxolane  $1^{4-7}$  or oxathiolane  $2^{8-10}$  have demonstrated the ability of these analogs to inhibit HIV replication  $^{6,9,10}$ .

Recently we have synthesized two series of dioxolane nucleoside homologs 3 and  $4^{11,12}$ . The most laborious step in their preparation was the separation of cis and trans isomers. To overcome this we decided to synthesize chiral nuc-

<sup>&</sup>lt;sup>2</sup> Semyonov Institute of Chemical Physics, the Russian Academy of Sciences, Kosygin str. 4, Moscow 117977, Russia;

<sup>&</sup>lt;sup>3</sup> Department of Pediatrics of the University of Alabama, 401 Volker Hall 1670, University Blvd., Birmingham, Alabama, 35294, USA.

Dedicated to the memory of Professor R.K.Robins.

616 MIKHAILOV ET AL.

leoside analogues on the basis of 4(R), 5(R)-dihydroxymethyl-2-methyl-1, 3-dioxolane. Optically active tartaric acids and their derivatives with the symmetry axis of second order  $C_2$  are frequently used in organic synthesis for similar purposes and this property may be transferred to their dioxolane derivatives too. It should be mentioned that 11 may be considered as analogues of 2',3'-dideoxy-3'-C-hydroxymethylnucleosides (5), exhibiting anti-HIV activity 14,15.

### RESULTS AND DISCUSSION

Starting from D-tartaric acid the known  $6^{16}$  was prepared and then transformed to  $7^{18}$ . 1,4-0-Dibenzyl-D-threitol  $^{18}$  was prepared in a good overall yield. Acid-catalyzed transacetalization of bromoacetaldehyde diethylacetal gave 9 in 74% yield. An attempt to use 1,4-0-dibenzoyl-D-threitol in this reaction failed due to migration of 0-benzoyl groups and formation of a complex reaction mixture.

Alkylation of the sodium salt of thymine with bromide 9 gave  $N^1, N^3$ -bis-alkylated and  $N^1$ -alkylated bases as main products  $^{12}$ . With the sodium salt of adenine the alkylation took place at  $N^9$  and  $N^3$  positions  $^{11}$ . Reaction of  $N^2$ -palmitoyl-guanine with 9 in the presence of potassium carbonate gave  $N^9$  and  $N^7$  isomers  $^{19}$ . The regio isomers were separated by adsorption chromatography on silica gel.

The benzyl protecting groups in 10 were removed by treatment with an excess of 10% palladium on carbon and ammonium formate in boiling methanol  $^{20}$ . For 10c, the palmitoyl group was removed by treatment with methanolic ammonia. The yields of 4(R), 5(R)-dihydroxymethyl-2-methyl-1, 3-dioxolane nucleosides 11 were moderate.

The structure of the compounds prepared was verified by  $^1\mathrm{H}$  NMR spectroscopy (tables 1 and 2). Rather complicated  $^1\mathrm{H}$  NMR spectra of 10 and 11 were analyzed using the reported data for 2,4-disubstituted 1,3-dioxolane nucleosides 3,  $^4\mathrm{I}^{11}$ ,  $^{12}$ .

The signals of methylene groups H2'a,b, H4'a,b, H5'a,b and protons H4 and H5 were unambiguously assigned by the NOE difference spectroscopy. Saturation of H2 in 10a exerted a rather strong NOE on the neighbouring protons H2'a,2'b (total effect > 4%). The cis oriented H5'a,5'b (1.2%) and H4 (2.9%) were also influenced significantly, while the effect on the

HOCH<sub>2</sub> O B HOCH<sub>2</sub> O CH<sub>2</sub> HOCH<sub>2</sub> O HOCH<sub>2</sub> O HOCH<sub>2</sub> O B HOCH<sub>2</sub> O CH<sub>2</sub> HOCH<sub>2</sub> O HOCH<sub>2</sub> O HOCH<sub>2</sub> S 
$$\frac{1 \text{ X} = 0}{2 \text{ X} = \text{ S}}$$
  $\frac{1 \text{ A} = 0}{2 \text{ X} = \text{ S}}$   $\frac{1 \text{ B} = 0}{3}$   $\frac{1 \text{ HC} + 0 \text{ CH}}{4 \text{ B}}$   $\frac{1 \text{ HC} + 0 \text{ CH}}{4 \text{ B}}$   $\frac{1 \text{ HC} + 0 \text{ CH}}{4 \text{ CH}}$   $\frac{1 \text{ A} + 0 \text{ CH}}{4 \text{ CH}}$   $\frac{1 \text{ A} + 0 \text{ CH}}{4 \text{ CH}}$   $\frac{1 \text{ A} + 0 \text{ CH}}{4 \text{ CH}}$   $\frac{1 \text{ A} + 0 \text{ CH}}{4 \text{ CH}}$   $\frac{1 \text{ A} + 0 \text{ CH}}{4 \text{ CH}}$   $\frac{1 \text{ A} + 0 \text{ CH}}{4 \text{ CH}}$   $\frac{1 \text{ A} + 0 \text{ CH}}{4 \text{ CH}}$   $\frac{1 \text{ C} + 0 \text{ CH}}{4 \text{ C} + 0 \text{ CH}}$   $\frac{1 \text{ C} + 0 \text{ CH}}{4 \text{ C} + 0 \text{ CH}}$   $\frac{1 \text{ C} + 0 \text{ CH}}{4 \text{ C} + 0 \text{ CH}}$   $\frac{1 \text{ C} + 0 \text{ CH}}{4 \text{ C} + 0 \text{ CH}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C} + 0 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C} + 0 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C} + 0 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C} + 0 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C} + 0 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C} + 0 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C} + 0 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C} + 0 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C} + 0 \text{ C}}{4 \text{ C}}$   $\frac{1 \text{ C}}{4 \text{ C}}$   $\frac$ 

B = a) Thy, b) Ade, c) Gua (in 10  $B = N^2-Pal-Gua$ )

a: EtOH/HCl,  $\Delta$ ; b: acetone/MeC(OEt)<sub>2</sub>Me/HCl; c: LiAlH<sub>4</sub>; d: 60% NaH, BzlCl/THF; e: 0,5 N HCl, MeOH,  $\Delta$ ; f: BrCH<sub>2</sub>CH(OEt)<sub>2</sub>/p-TsOH; g: sodium salts of thymine and adenine, N<sup>2</sup>-palmitoylguanine/K<sub>2</sub>CO<sub>3</sub>, DMF; h: 10% Pd/C, HCOONH<sub>4</sub>, MeOH.

**SCHEME** 

TABLE 1.  $^{1}$ H NMR chemical shifts ( $\delta$ ) for the dioxolane nucleoside analogues 10 in CDCl $_{3}$  and 11 in D $_{2}$ O at 300 K.

Compd. Base			Dioxolane moiety									
			Н2	H2′a	H2′b	Н4	H4'a	Н4′Ъ	Н5	H5′a	Н5′Ь	
10a	7.03 q	-	5.23		3.81 dd	4.08 ddd		3.47 dd			3.51 dd	
10b	8.30 s		5.37 t			4.08 ddd					3.47 dd	
10c	8.31 s		5.38 t						3.90 ddd	3.55 dd	3.51 dd	
11a	7.36 q					3.88 ddd						
11b	8.58 s	8.51 s	5.89 t	4.86 d	4.86 d	4.34 ddd	3.76 dd	3.59 dd	4.07		3.95	
11c	7.95 s		5.58 t		4.41 dd			3.43 dd			3.74 dd	

Benzyl group: 7.30m (10H, 2Ph), 4.48s ( $2H, CH_2$ ), 4.45s ( $2H, CH_2$ ) Palmitoyl group: 2.48t ( $2H, CH_2$ ), 1.70m ( $2H, CH_2$ ), 1.30brs ( $24H, CH_2$ ), 0.87t (3H, Me).

TABLE 2. Coupling constants (J in Hz) for 10 and 11 at 300 K.

2,2á	2,2b	2á,2b	4,4å	4,4b	4á,4b	4,5	5,5å	5,5b	5á,5b
3.5	3.7	-14.3	4.6	4.8	-10.3	6.3	4.6	5.1	-10.3
3.0	3.0	-14.4	5.0	5.0	-10.4	6.3	4.2	4.8	-10.4
2.5	2.5		4.3	5.5	-10.3	6.2	4.0	4.9	-10.5
2.9	2.9	-14.9	3.6	5.7	-12.3	6.6	3.5	5.7	-12.5
2.5	2.5	-	3.7	5.9	-12.1	6.4	-	_	-
2.6	2.6	-15.2	4.0	6.0	-12.1	6.4	4.7	5.9	-12.3
2	3.5 3.0 2.5 2.9 2.5	3.5 3.7 3.0 3.0 2.5 2.5 2.9 2.9 2.5 2.5	3.5 3.7 -14.3 3.0 3.0 -14.4 2.5 2.5 - 2.9 2.9 -14.9 2.5 2.5 -	3.5 3.7 -14.3 4.6 3.0 3.0 -14.4 5.0 2.5 2.5 - 4.3 2.9 2.9 -14.9 3.6 2.5 2.5 - 3.7	3.5 3.7 -14.3 4.6 4.8 3.0 3.0 -14.4 5.0 5.0 2.5 2.5 - 4.3 5.5 2.9 2.9 -14.9 3.6 5.7 2.5 2.5 - 3.7 5.9	3.5 3.7 -14.3 4.6 4.8 -10.3 3.0 3.0 -14.4 5.0 5.0 -10.4 3.5 2.5 - 4.3 5.5 -10.3 3.9 2.9 -14.9 3.6 5.7 -12.3 3.5 2.5 - 3.7 5.9 -12.1	3.5 3.7 -14.3 4.6 4.8 -10.3 6.3 3.0 3.0 -14.4 5.0 5.0 -10.4 6.3 3.5 2.5 - 4.3 5.5 -10.3 6.2 3.9 2.9 -14.9 3.6 5.7 -12.3 6.6 3.5 2.5 - 3.7 5.9 -12.1 6.4	3.5 3.7 -14.3 4.6 4.8 -10.3 6.3 4.6 3.0 3.0 -14.4 5.0 5.0 -10.4 6.3 4.2 3.5 2.5 - 4.3 5.5 -10.3 6.2 4.0 3.9 2.9 -14.9 3.6 5.7 -12.3 6.6 3.5 3.5 2.5 - 3.7 5.9 -12.1 6.4 -	2,2á       2,2b       2á,2b       4,4á       4,4b       4á,4b       4,5       5,5á       5,5b         3,5       3,7       -14,3       4,6       4,8       -10,3       6,3       4,6       5,1         3,0       3,0       -14,4       5,0       5,0       -10,4       6,3       4,2       4,8         2,5       2,5       -       4,3       5,5       -10,3       6,2       4,0       4,9         2,9       2,9       -14,9       3,6       5,7       -12,3       6,6       3,5       5,7         2,5       2,5       -       3,7       5,9       -12,1       6,4       -       -         2,6       2,6       -15,2       4,0       6,0       -12,1       6,4       4,7       5,9

J<sub>5,6</sub> 1.2 (Thy).

trans oriented 4'a,4'b and H5 remained near zero. The assignment in the other derivatives 10b and 10c was carried out similarly.

It should be noted that the geminal constant  $J_{2'a,2'b}$  in  $CH_2N$  group is greater than that in  $CH_2O$  group  $(J_{4'a,4'b}$  and  $J_{5'a,5'b})$  (tabl. 2). The vicinal coupling constants  $J_{4,5}$  in 10 and 11 were in the range of 6.2-6.6 Hz.

The deprotected nucleoside analogues 11 were UV spectroscopically almost identical with their parent nucleosides thymidine, adenosine and guanosine. A low negative Cotton effect with  $\lambda_{max}$  268 nm ( $\Delta\epsilon$  -0.22) was observed in CD spectra of thymine nucleoside 11a.

The above described nucleoside analogues were found to be inactive against HIV-1 (CEM cells) and HSV-1, HSV-2, HCMV, VZV (HFF cells) at concentration up to 100  $\mu$ g/ml and were nontoxic towards CEM and HFF cells. The absence of activity is probably due to the fact that 4(R), 5(R)-dihydroxymethyl-2-methyl-1,3-dioxolane structures are not being recognized by selective cellular and viral kinases.

#### **EXPERIMENTAL**

General methods. Melting points (uncorrected) were determined with a TP (USSR) instrument. UV spectra were recorded on a Specord UV-vis spectrometer. Silica gel L (40-100  $\mu$ m) (Czechoslovakia) was used for adsorption column chromatography. TLC was carried out on Silufol UV<sub>254</sub> (Kavalier, Czechoslovakia) and Kieselgel 60 F<sub>254</sub> (Merck, Germany) using A CHCl<sub>3</sub>, B 95:5 CHCl<sub>3</sub>-EtOH, C 9:1 CHCl<sub>3</sub>-EtOH, and D 8:2 CHCl<sub>3</sub>-EtOH and detection by UV light. All crystalline compounds gave correct elemental analyses ( $^{\pm}$ 0.4%).

<sup>1</sup>H NMR spectra were recorded using a Bruker AMX 400 spectrometer at 300 K. Chemical shifts were measured relative to solvent signals. The signals were assigned by the double resonance techniques. The NOE measurements in CDCl<sub>3</sub> were performed under identical spectral and processing conditions by applying an NOEDIFF pulse sequence of Bruker software package UXNMR (release version 911101) for steady-state NOE measurements. For compounds 10 and 11, the values of chemical shifts and coupling constants were calculated by means of a DAISY programme on a Aspect X32/2 computer.

620 MIKHAILOV ET AL.

2-Bromomethy l-4(R), S(R)-dibenzy loxymethy l-1, 3-dioxolane (9). A mixture of 1,4-di-O-benzyl-D-threitol 5 (6.1 g, 20.2 mmol), bromoacetal dehyde diethyl acetal (2.42 mL, 20.2 mmol) and p-toluenesul fonic acid (0.2 g) in dry acetonitrile (20 mL) was gently heated for 2 h and evaporated in vacuo to dryness. The residue was diluted with chloroform (150 mL), washed successively with saturated aqueous sodium hydrogencarbonate (30 mL) and water (2x30 mL), dried with Na<sub>2</sub>SO<sub>4</sub>, filtered and evaporated in vacuo to dryness. The residue was chromatographed on silica gel (50 g), using a chloroform-hexane (1:4 v/v) mixture. The pooled fractions were evaporated to a syrup. Yield 6.1 g (74%).  $R_F$  0.93 (A). <sup>1</sup>H NMR (CDCl<sub>3</sub>): 7.26 brs (10H, 2CH<sub>2</sub>Ph), 5.27 t (1H, J 4.0, CHCH<sub>2</sub>), 4.55 brs (2H, CH<sub>2</sub>Ph), 4.54 brs (2H, CH<sub>2</sub>Ph), 4.20-4.08 m (2H, 2CH), 3.66-3.56 m (4H, 2CH<sub>2</sub>), 3.39 d (2H, J 4.0, CHCH<sub>2</sub>).

4(R), 5(R)-Dibenzy loxymethy l-2-(thymin-1-ylmethy 1)-1, 3dioxolane (10a). To a suspension of dry thymine (1.04 g, 8.25 mmol) in dry dimethylformamide (DMF, 30 mL) was added sodium hydride (0.4 g, 10.3 mmol, 60% in oil) and the mixture was stirred for 1 h at  $20^{\circ}$ C. Then the mixture was heated to  $110^{\circ}$ C and a solution of 9 (3.05 g, 7.5 mmol) in DMF (10 mL) was added in several portions over 2 h. The mixture was heated over 10 h, cooled to 20°C, filtered and the combined filtrate and DMF washings were evaporated in vacuo to dryness. The residue was dissolved in chloroform (150 mL), the organic layer was washed with water (2x30 mL), dried with Na<sub>2</sub>SO<sub>4</sub>, filtered, evaporated to dryness and chromatographed on silica gel (100 g). Elution with a chloroform-hexane (1:1 v/v) mixture gave the starting bromide 8 and with system A gave  $N^{1}$ ,  $N^{3}$ -bis-product (yield 0.32 g, 5%). Further elution with system A gave 10a as a syrup. Yield 0.37 g (10%).  $R_F$  0.42 (B). The  $^{1}$ H NMR data are given in Tables 1 and 2.

4(R), 5(R)-Dibenzyloxymethyl-2-(adenin-9-ylmethyl)-1,3-dioxolane (10b). This was prepared analogously by alkylation of sodium salt of adenine (4.73 mmol) with 9 (1.75 g, 4.3 mmol) in dry DMF (15 mL). The products were separated on silica gel (system B) to give 10b. Yield 0.9 g (41%).  $R_F$  0.57 (B). M.p. 117-119<sup>OC</sup> (ethanol). Further elution with the same solvent gave the corresponding  $N^3$ -isomer. Yield 0.20 g (4%).  $R_F$  0.35 (B).

4(R), 5(R) - Diberzy loxymethy  $l-2-(N^2-palmitoy lguan in-9-y l$ methyl)-1,3-dioxolane (10c). To a stirred suspension of  $N^2$ palmitoylguanine (1.45 g, 3.72 mmol) and  $K_2CO_3$  (0.75 g, 5.4 mmol) in dry DMF (200 mL) a solution of 9 ( $\overline{3}$  g, 7.37 mmol) in dry DMF (20 mL) was added in several portions over 4 h at 20°C. The mixture was stirred for 3 days at 20°C and evaporated in vacuo to dryness. The residue was dissolved in chloroform (300 mL), the organic layer was washed with water (2x50 mL), dried with Na<sub>2</sub>SO<sub>4</sub>, filtered through Hyflo Super Cel, evaporated to dryness and chromatographed on silica gel (100 g). A column was washed with system A (400 mL), further elution with a chloroform-ethanol (98.5:1.5 v/v) mixture gave N-7 isomer. Yield 0.35 g (13%).  $R_F$  0.60 (B). Further elution with the same mixture gave 10c as a syrup. Yield 0.35 g (13%).  $R_F$  0.47 (B).

4(R), 5(R)-Dihydroxymethy l-2-(thymin-1-ylmethy l)-1, 3-dio-xolane (11a). To a solution of 10a (0.2 g, 0.44 mmol) in methanol (20 mL) HCOONH<sub>4</sub> (250 mg) and 10% Pd/C (Fluka, 154 mg, 350 mg for 1 mmol) were added, the mixture was refluxed for 3 h, filtered through Hyflo Super Cel and the combined filtrate and methanol washings were evaporated to dryness. The residue was dissolved in water (70 mL), the aqueous layer was washed with ethyl acetate (2x20 mL) and evaporated to dryness. Product 11a slowly crystallized from ethanol. Yield 78 mg (66%).  $R_F$  0.23 (B). M.p. 135-136°C. UV:  $\lambda_{max}^{pH1-7}$  271 nm ( $\epsilon$  8700);  $\lambda_{max}^{pH13}$  271 nm ( $\epsilon$  6500).

4(R), 5(R)-Dihydroxymethyl-2-(adenin-9-ylmethyl)-1, 3-dio-xolane (11b). This was prepared analogously starting from 10b (0.2 g, 0.43 mmol). Product 11b slowly crystallized from acetone. Yield 90 mg (70%). R<sub>F</sub> 0.10 (B). M.p. 145-147°C. UV:  $\lambda_{\text{max}}^{\text{pH1}}$  258 nm ( $\epsilon$  12520);  $\lambda_{\text{max}}^{\text{pH7}-13}$  261 nm ( $\epsilon$  12860).

4(R), 5(R)-Dihydroxymethyl-2-(guanin-9-ylmethyl)-1, 3-dio-xolane (11c). A mixture of 10c (0.32 g, 0.45 mmol), HCOONH<sub>4</sub> (250 mg) and 10% Pd/C (154 mg) in methanol (20 mL) was refluxed for 2 days, filtered through Hyflo Super Cel and the combined filtrate and methanol washings were evaporated to dryness. The residue was dissolved in methanolic 5M ammonia (20 mL), kept for 3 days at  $20^{\circ}$ C and then concentrated to dry-

622 MIKHAILOV ET AL.

ness. The residue was partitioned between water (50 mL) and chloroform (20 mL), and the organic layer was washed with water (20 mL). The combined aqueous extracts were washed with chloroform (20 mL), concentrated to dryness, and the residue was recrystallized from ethanol. Yield 53 mg (40%).  $R_F$  0.08 (B). M.p. 187-189°C. UV:  $\lambda_{\text{max}}^{\text{pH-1}}$  258 nm (\$\varepsilon\$ 10790);  $\lambda_{\text{max}}^{\text{pH-7}}$  252 nm (\$\varepsilon\$ 11090);  $\lambda_{\text{max}}^{\text{pH-13}}$  268 nm (\$\varepsilon\$ 9500).

#### **ACKNOWLEDGEMENTS**

This research was supported by the Russian Fund of Fundamental Sciences, project 93-04-7607(S.N.M.) and NIH, NIAD grant No NO1 AI 82518 (E.R.K.). We would like to thank "Bruker Analytische Messtechnick GmbH" (Karlsruhe, Germany) for providing an opportunity to perform our measurements on NMR spectrometer AMX400 in the Demo Center in Moscow. We are greatful to Dr. C.Tseng from the National Institute of Allergy and Infectious Diseases (NIAID) and Dr. R.Schultz from the National Cancer Institute (NCI) for the antiviral screening.

#### REFERENCES

- Mitsuya H., Weinhold K.J., Furman P.A., St.Clair M.H., Lerman N.S., Gallo R.G., Bolognesi D., Barry D.W., Broder S. Proc. Natl. Acad. Sci. USA, 1985, 82, 7096.
- Mitsuya H., Broder S. Proc. Natl. Acad. Sci. USA, 1986, 83, 1911.
- 3. Design of Anti-AIDS Drug. (Edited by E. De Clercq).
  Amsterdam, Elsevier, Pharmac. Library, 1990, 14, 372p.
- Wainberg M.A., Stern M., Martel R., Belleau B., Soudeyns H. Abstr. Papers, V<sup>th</sup> Inter. Confer. on AIDS, Montreal, Ontario, 1989, M.C.P. 63.
- 5. Norbeck D.W., Spanton S., Broder S., Mitsuya H. Tetrahedron Lett., 1989, 30, 6263.
- Kim H.O., Ahn S.K., Alves A.J., Beach W.J., Jeong L.S., Choi B.G., Van Roey P., Schinazi R.F., Chu C.K. J. Med. Chem., 1992, 35, 1987.
- 7. Kim H.O., Schinazi R.F., Shanmuganathan K., Jeong L.S., Beach J.W., Nampalli S., Cannon D.L., Chu C.K. J. Med. Chem., 1993, 36, 519.
- 8. Belleau B., Dixit D., Nguyen-Ba N., Kraus J-L. Abstr. Papers, V<sup>th</sup> Inter. Confer. on AIDS, Montreal, Ontario, 1989, T.C.O.1.

- Beach J.W., Jeong L.S., Alves A.J., Pohl D., Kim H.O., Chang C-N., Doong S-L., Schinazi R.F., Cheng Y-C., Chu C.K. J. Org. Chem., 1992, 57, 2217.
- Jeong L.S., Schinazi R.F., Beach J.W., Kim H.O., Nampalli S., Shanmuganathan K., Alves A.J., McMillan A., Chu C.K., Mathis R. J. Med. Chem., 1993, 36, 181.
- 11. Efimtseva E.V., Mikhailov S.N., Meshkov S.V., Lonnberg H. Acta Chem. Scand. 1992, 46, 1122.
- 12. Efimtseva E.V., Mikhailov S.N., Meshkov S.V., Bochkarev A.V., Gurskaya G.V. Bioorg. Khim. (Russia), 1993, 19, 103.
- Mash E.A. Homochiral ketals and acetals in organic synthesis. In: Studies in Natural Products Chemistry, 1988, p. 577-653.
- Tseng C.K.-H., Marques V.E., Milne G.W., Wisocki R.J.Jr., Mitsuya H., Shirasaki T., Driscoll J.S. J. Med.Chem., 1991, 34, 343.
- Svansson L., Kvarnstrom I., Classon B., Samuelsson B.
   Org. Chem., 1991, 56, 2993.
- 16. Kuhn P., Wagner-Jauregg O. Chem. Ber., 1928, 61, 483.
- 17. Holy A. Collect. Czech. Chem. Commun., 1979, 44, 593.
- 18. Ando N., Yamamoto Y., Oda J., Inouye Y. Synthesis, 1978, 9, 688.
- 19. Kjellberg J., Johansson N.G. Nucleos. Nucleot. 1989, 8, 225.
- 20. Bieg T., Szeja W. Synthesis, 1985, N 1, 76.

Received 8/17/93 Accepted 10/21/93